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Synthesis of Magnetic Activated Carbon Treated with Sodium Dodecyl Sulphate

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ABSTRACT

Magnetic activated carbon (MAC) is found to be effective for the adsorption of methylene blue due to its physico-chemical properties such as strong adsorption of magnetization. The use of activated carbon (AC) for methylene blue adsorption was ineffective compared to MAC. MAC was prepared by incorporating different types of iron powder and chemicals [sodium dodecyl sulphate (SDS), citric acid (CA), dimethicone (D350), and epichlorohydrin (C_3H_5ClO)] to strengthen the magnetism and stabilize the MAC. The methylene blue test and iodine test were tested on different samples. Characterization test on physical and chemical properties was carried out using Scanning Electron Microscopy (SEM) and X-ray diffraction (XRD). The yield of MAC was higher because of the addition of magnetic particles. The incorporation of magnetic particles had been proven by the SEM and XRD analysis that showed the presence of iron compound. The performance study of the adsorbent sample showed that MAC A3II presented better qualities with highest removal percentage (98.81 % of removal) in methylene blue adsorption and low magnetic contact time that showed strong magnetism. MAC A3II was prepared by incorporating iron powder and treated by using sodium dodecyl sulphate (SDS). Among all the adsorbent sample, MAC B2III performed the weakest quality because the dye removal percentage was low, and the preparation process was complicated compared with others.

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INTRODUCTION

Batik is a traditional fabric craft with handpainted and richly colored patterns. The batik industry is one of economic driven industries in east coast states of Malaysia- Kelantan and Terengganu (Jamaludin, 2020). Many

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batik entrepreneurs built their manufacturing units along the adjoining riverside for disposal of wastewater such as dyes, grease, wax, and heavy metals (Ramakreshnan et al., 2020). Methylene blue (MB) is commonly used dye in batik industry. The presence of dye in rivers not only deteriorates aesthetic value of water but also compromises human health (Ewadh, 2020). Thus, a proper treatment for effluents rich in color and organic pollutants is required prior to discharging into the environment (Ramakreshnan et al., 2020). One of the methods applied to treat the wastewater is Advanced Oxidation Processes (AOP). In this method, the spent bleaching earth impregnated with Tungsten trioxide went through sonocatalytic to form hydroxyl radicals. The hydroxyl radicals therefore destruct pollution compounds. Although the percentage of pollutant degradation is 99.12% the process is expensive for a small cottage batik manufacture (Hindryawati et al., 2020). Therefore, adsorption is an effective method to treat wastewater. Many industries used commercial AC to treat effluents from factories, the drawbacks of AC are high cost and disposal problem (Vyavahare et al., 2019). Thus, current research found the potential of agricultural waste to produce activated carbon. By this way, it is not only cost effective but also environment- friendly (Shokry et al., 2020). Furthermore, production on magnetic activated carbon (MAC) from agricultural waste represents novel adsorbent for water purification. Activated carbon is a high-content carbon enriched with minerals adsorbent. By this way, it is not only cost effective but also environment- friendly (Shokry et al., 2020) Activated carbon can be produced through the pyrolysis of biomass in the absence of oxygen at \leq 500°C or carbonization process and it is a carbon rich residue. Activation step of activated carbon involved oxidizing gases such as steam, air, and carbon dioxide (Ooi et al., 2013). It has porous properties which can increase the removal of contaminants from aqueous solution. It also has large specific surface area and strong redox reactivity (Yao et al., 2015). Therefore, unique properties of activated carbon are widely used as a filter to improve the quality of water and wastewater whereby it increases efficiency of water treatment and reduces the spread of contamination from hazardous chemicals. Accordingly, MAC is proposed to improve effectiveness of separation process which aided by external magnetic field (Shokry et al., 2020). The key obstacle of impregnation process is magnetic particles does not fully impregnate with the AC. In order to encounter the problem, a surfactant is applied due to change the surface properties of adsorbent. The hydrophobic and hydrophilic characteristics of the surfactant enable improve adsorption capacity by affecting solubility of the organic pollutant. Thus, organic pollutants such as dye is suggested to test the performance of impregnation process with surfactant treatment (Li et al., 2019). An alternative method to produce magnetic activated carbon derived from coconut shell with sodium dodecyl sulfate treatment is proposed in this study. Besides, the methylene blue will be used as pollutant to test the adsorbent. The challenge of this study is to show that MAC is a competitive adsorbent by comparing with commercial AC. Hence, the objective of this study was to synthesis magnetic activated carbon treated with SDS and to test the performance of the novel adsorbent.

MATERIALS AND METHOD

Chemical and Reagents

The chemical and reagent used in the research were hydrochloric acid, HCl (37%), ammonium hydroxide, NH₄OH (28%), 2 M hydrochloric acid, HCl (2 M), ammonium hydroxide, NH₄OH (0.7 M), iodine solution (0.1 N), sodium thiosulphate solution, Na₂S₂O₃ (0.05 N), starch solution, sodium carbonate, Na₂CO₃, potassium iodide (KI), and distilled water. The magnetization of AC was carried out using iron powder (IFP) and labelled as Group A; Group B was γ -Fe₂O₃ (I30B), the gamma phase of ferrous oxide; Group C was α -Fe₂O₃ (I30) the alpha phase of ferrous oxide; Group D was ferrous chloride, FeCl₂ impregnated with ferric chloride, FeCl₃ and Group E was ferrous sulphate, FeSO₄ impregnate with Ferric Chloride, FeCl₃. In addition, sodium dodecyl sulphate (SDS), citric acid (CA), dimethicone 350 (D350) and epichlorohydrin, (C₃H₅ClO) used to strengthen the impregnation between magnetic particles and AC.

Synthesis of Activated Carbon

Raw coconut shell sample was collected from coconut milk manufacturer company, which is located at Tanah Merah district, Kelantan state. The raw coconut shell was carbonized by using modified drum method. Carbonization is a conversion of organic matters like biomass into carbon through destructive distillation. The activated carbon was produced through modified pyrolysis technique using an empty 55-gallon steel oil drum. The coconut shell was placed into the oil drum and lit from the bottom. Once the materials inside the drum were fully ignited and the water in the carbon source had evaporated, the drum was sealed to initiate the anoxic combustion process. The process would take about two to three hours for the material to be fully carbonized. Once the charcoal was completed, it was sorted by visual inspection to find material that had been thoroughly carbonized. Shells that retained a natural brown colour and that were difficult to break by hand were excluded. Any material that had not been completely carbonized was saved for the following batch. The drum method produced approximately 5 kg of carbon material. There is no fixed temperature in this process but can ranged from 400 to 500°C (Cobb et al., 2012). The schematic diagram on the preparation of activated carbon using carbonization drum method is illustrated in Figure 1. The carbonized coconut shells were grounded into the desired form and further subjected to activation with 1N concentration of potassium hydroxide (KOH) at a ratio of 1:3, with slow agitation at 75 rpm for 1 hour. To prepare 1N KOH, 56.11 g of KOH was dissolved in 300 mL water and then transferred to a 1 L measuring flask quantitatively and dilute until it reaches 1 liter (Cobb et al., 2012). The mixture was left to mature for about 5 to 6 hour and followed by filtration, later, rinsed with double distilled water (ddH_2O). The AC was dried in oven at temperature of 90 -100°C. The dried AC was placed in a muffle furnace (Carbolite ELF 11/6B) and heated to 800-900°C (10°C/min), and kept for 15-30

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Figure 1. Schematic diagram on the synthesis of activated carbon from coconut shell via carbonization method

min (Lee et al., 2018). The cooled down sample was washed, neutralized with 5% HCl, dried and stored for further modification.

Preparation Method for Magnetization of Coconut Shell Activated Carbon

Table 1 displays the different combination of the final product by using different magnetic particles impregnated with different chemical and different sonicated treatment. The AC used was in mesh size of less than 45 μ m. At first, 2.95 g of iron powder or iron (III) oxide powder, Fe₂O₃ (IFP, I30B, and I30) was dissolved in 45 mL of distilled water and placed in the orbital shaker for the agitation of 250 rpm for 30 min. For MAC, the experiment was continued with the addition of 6 mL of ammonium hydroxide solution, NH₄OH (28%) and agitated for 1 hour. Next, 1.5 g of AC was added and continued the agitation for 5 min. Then, the preparation was continued with addition of 0.6 ml of different chemical (control, Dimetricone 350, Sodium Dodecyl Sulphate, and Citric Acid) respectively as enhancer of magnetization and continue agitated for another 1 hour. All of the samples were continued with 3 different sonication for 1 hour which is non-sonicated, ultrasonic bath and sonicated respectively. Lastly, the final product, MAC was washed and collected using neodymium magnet. The MAC was dried at 50°C in the oven to obtain the dry MAC powder. Group C (I30) contained the lowest magnetism among all the group which the magnetic particles used was the alpha phase of ferric oxide, α -Fe₂O₃ which was the lowest magnetism among

all magnetic particles. D350 was the worst chemical used to treat the sample which wasted lots of carbon sample during the washing process.

		Non- sonicated (I)	Ultrasonic bath (II)	Sonicated (III)
	Control (1)	MAC_A1I	MAC_A1II	MAC_A1III
IED (A)	D350 (2)	MAC_A2I	MAC_A2II	MAC_A2III
IFF (A)	SDS (3)	MAC_A3I	MAC_A3II	MAC_A3III
	CA (4)	MAC_A4I	MAC_A4II	MAC_A4III
	Control (1)	MAC_B1I	MAC_B1II	MAC_B1III
120D (D)	D350 (2)	MAC_B2I	MAC_B2II	MAC_B2III
130Б (Б)	SDS (3)	MAC_B3I	MAC_B3II	MAC_B3III
	CA (4)	MAC_B4I	MAC_B4II	MAC_B4III
	Control (1)	MAC_C1I	MAC_C1II	MAC_C1III
120(C)	D350 (2)	MAC_C2I	MAC_C2II	MAC_C2III
150 (C)	SDS (3)	MAC_C3I	MAC_C3II	MAC_C3III
	CA (4)	MAC_C4I	MAC_C4II	MAC_C4III

Sample with different treatment combination using method 1

Table 1

Table 2

Notes: A = IFP (Iron Powder); B = I30B (γ -Fe₂O₃); C = I30 (α -Fe₂O₃); 1 = Control; 2 = D350; 3 = SDS; 4 = CA; I = Non-sonicated; II = Ultrasonic bath; III = Sonicated

Preparation Method for Magnetization of Coconut Shell Activated Carbon using Method 2

For the impregnation magnetization 1.95 g of FeCl₃ was mixed with 0.7 g of FeCl₂ (D) and then dissolved in 45 mL of distilled water and placed in the orbital shaker for the agitation at 250 rpm for 30 min (Table 2). The sample was further added with 6 mL of ammonium hydroxide solution, NH₄OH (28%) and continued agitation for another 1 hour. Then, 1.5 g of AC was added into the solution and the agitation continued for another 5 mins. The prepared solution was added with different chemical (control, SDS, and CA) and continued the agitation for another 1 hour. The sample were treated using three different method for 1 hour which was non-sonicated, ultrasonic bath and sonicated respectively. Lastly, the

		Non- sonicated (I)	Ultrasonic bath (II)
$FeCl_3 + FeCl_3 (D)$	Control (1)	MAC_D1I	MAC_D1II
	SDS (3)	MAC_D3I	MAC_D3II
	CA (4)	MAC_D4I	MAC_D4II
$FeCl_3 + FeSO_4 (E)$	Control (1)	MAC_E1I	MAC_E1II
	SDS (3)	MAC_E3I	MAC_E3II
	CA (4)	MAC_E4I	MAC_E4II

Sample with different treatment combination using method 2

Notes: D = FeCl₂; E = FeSO₄; 1 = Control; 3 = SDS; 4 = CA; I = Non-sonicated; II = Ultrasonic bath

final product, was washed and collected using neodymium magnet. The treated MAC was dried at 50 °C in the oven to obtain the dry MAC powder. The magnetization method was repeated $FeCl_3 + FeSO_4(E)$ by replacing the preparation formula (i) with formula (ii) (1.95 g of $FeCl_3$ impregnated with 1.0 g of $FeSO_4$).

Process Performance

Yield (%). The total yield of MAC (%) estimated by using the following Equation 1,

Total Yield of MAC (%) =
$$\frac{\text{weight of MAC}_XX \text{ produced}}{\text{weight of activated carbon sample}} \times 100\%$$
 (1)

Methylene Blue Test. Methylene blue test was carried out using method described by Jadhav and Mohanraj (2016). Methylene blue solution was prepared by diluting the stock solution with 1000 ppm to 2 ppm. A fixed parameter was measured in this adsorption experiment where the time taken of the MAC to decolorize the methylene blue (contact time) and the time taken of the sample fully attached to the magnet (magnetic contact time) were recorded. At first, 0.15 g of each sample were weighed and poured into 25 mL of methylene blue solution. The time taken to decolorize the methylene blue and the time of the sample fully attached to the magnet were recorded. The methylene blue test was conducted in triplicates under identical condition and average value was calculated. Next, the filtrate was analyzed using UV visible spectrophotometer (Thermo Scientific Genesys 20) at 668 nm (Islam et al., 2017). This was to further prove the effectiveness of the MAC in adsorption of methylene blue solution. The absorbance value was recorded in triplicate and the average value were also recorded. The percentage of dye removal by using different sample was calculated using mathematical Equation 2.

Percentage of dye removal (%) =
$$\frac{(B-S)}{S} \times 100$$
 (2)

Where B (mg/ L) and S (mg/ L) are MB concentration at initial and after added MAC. The steps were repeated by using different concentration of methylene blue dye which was 5 ppm and 10 ppm.

Iodine Test. Iodine number was used to determine the adsorption of iodine on the activated carbon (Mianowski et al., 2007). The main principle was to determine the residual iodine concentration by titration with sodium thiosulfate standard solution after the quantitative sample and iodine standard solution were fully shaken under specific conditions to determine the iodine adsorption amount. Iodine adsorption value can be used as a qualitative indicator of microporous structure of activated carbon (Yang et al., 2020).

Preparation of Iodine Solution (0.1 N). For the preparation of iodine solution (0.1 N), 12.7 g of iodine pearls and 19.1 g of potassium iodide (KI) were weighed. Next, around 2-5 mL of distilled water was added slowly and stirred using magnetic stirrer. Distilled water was added until 50 mL to completely dissolve the iodine pearl. The solution was allowed to stir for a minimum of 4 hours to make sure that all the crystals were completely dissolved. The solution was quantitatively transferred to a 1 L volumetric flask (ASTMD4607_14, 2014). The iodine solution 0.1 N must be stored in amber bottle due to its sensitivity to light.

Preparation of Sodium Thiosulphate, $Na_2S_2O_3$ (0.05 N). Around 12.4 g of sodium thiosulphate, $Na_2S_2O_3$ (0.05N) was dissolved in approximately 100 mL of freshly boiled distilled water and 0.1 g of sodium bicarbonate, NaHCO₃ was added to decrease the bacterial decomposition of the thiosulphate solution. The mixture was transferred quantitatively to a 1 L volumetric flask and diluted to the mark (ASTMD460714, 2014).

Preparation of Starch Solution. To prepare a starch solution, 1.0 ± 0.5 g of starch was mixed with 5 to 10 mL of cold water to make a paste. Additional of 25 ± 5 mL of water was added while stirring to the starch paste. The mixture was poured into a 1 L boiling water and boiled for 4 to 5 min (ASTMD460714, 2014).

Iodine Number. Iodine number was determined on iodine solution as blank, 45 µm of AC, 63 µm of AC, CMAC, the best and worst condition of sample, MAC XX. The test divided into two parts as part a and b. Part b was a blank reading. At first, 10 mL of iodine solution (0.1 N) was measured and poured into conical flask. The solution was titrated with sodium thiosulphate solution, (0.05 N) until the solution turn to pale yellow. Then, 2 drops of starch solution were added, and the solution turned blue. The solution was continued with titration of sodium thiosulphate until the solution turned colorless. The burette reading corresponds to blank reading (b). For part a, around 0.2 g of sample (AC, MAC, MAC XX) was weighed and dissolved in 40 mL of iodine solution 0.1N. The solution was shaken for 50 min at 200 rpm in the orbital shaker and filtered. Then, 10 mL of filtrate was measured and poured into another conical flask. The filtrate was titrated with sodium thiosulphate solution until the solution turns pale yellow. Next, 2 drops of starch solution were added into the solution until the solution turns blue. The solution was continued with the titration of sodium thiosulphate until the solution turned colorless. The burette readings were correspond to (a). The iodine value was calculated by using Equations 3 to 5 as stated below.

Indine value = $c \times Conversion factor; mg/g$ (3)

Conversion factor =

molecular weight of iodine \times normality of iodine \times 40 \times blank re	adina
weight of activated carbon	aung
$=\frac{127\times0.1\text{ N}\times40}{0.2\text{ g}}\times\text{b}$	(4)
c = b - a	(5)

Characterization of magnetic activated carbon

Scanning Electron Microscopy (SEM) Analysis. SEM was used to analyze the pore structure of AC and MAC sample that produced. The magnification used were 1000×. The sample was placed on the SEM holder and moulded into the mounting holed in a JEOL JSM-IT100 scanning electron microscope. The focus knob was adjusted and the selected image captured.

X-ray Diffraction (XRD). X-ray diffraction is method a that uses X-ray to characterize the crystalline properties of carbon material. The sample was packed into the sample holder and analyzed by using Bruker D2 Phaser. The sample was scanned from 20.0°C to 90.0°C at the scan speed of 2° min⁻¹. The XRD diffractogram patterns was obtained from the system.

RESULTS AND DISCUSSION

Weight and Yield of Magnetic Activated Carbon

Table 3 shows the weight and the total yield of the MAC. Magnetic activated carbon was produced by using different type of iron powder (A, B, C, D, and E), different chemical to treat MAC (1, 2, 3, and 4) and also different type of sonication (I, II, and III). The production yield was considered as an important parameter in the feasibility study under specified condition. The magnetism of these MAC sample was tested using neodymium magnet (NdFeB). The results showed that the magnetism of group C was the lowest. The results clearly showed that the MAC yield of all samples was more than 100 %. This is because the addition of different type of iron powder into different samples. The highest MAC yield, 417.33% was achieved by MAC_A4II (True yield: 140.67%). The lowest MAC yield among all sample was from MAC_E3II that recorded 86% (True yield: 28.99%). This happened because of the washing and cleaning process that washed away most of the AC. Besides, this could also be due to the failure of impregnation process to impregnate the FeSO₄ powder and the AC.

No	Sample	Weight (g)	Total Yield of MAC (%)	True Yield (%)
1	MAC A1I	3.29	219.33	73.93
2	MAC A1II	4.74	316.00	106.52
3	MAC A2I	3.34	222.67	75.06
4	MAC A2II	3.85	256.67	86.52
5	MAC_A2III	3.28	218.67	73.71
6	MAC A3I	4.80	320.00	107.87
7	MAC_A3II	4.94	329.33	111.01
8	MAC_A3III	4.66	310.67	104.72
9	MAC_A4I	4.61	307.33	103.60
10	MAC_A4II	6.26	417.33	140.67
11	MAC_A4III	4.16	277.33	93.48
1	MAC_B1I	4.14	276.00	93.03
2	MAC_B1II	4.31	287.33	96.85
3	MAC_B2I	3.71	247.33	83.37
4	MAC_B2II	3.53	235.33	79.33
5	MAC_B2III	3.82	254.67	85.84
6	MAC_B3I	3.50	233.33	78.65
7	MAC_B3II	3.61	240.67	81.12
8	MAC_B3III	3.93	262.00	88.31
9	MAC_B4I	3.12	208.00	70.11
10	MAC_B4II	4.71	314.00	105.84
11	MAC_B4III	3.61	240.67	81.12
12	MAC_C1I	4.08	272.00	91.69
13	MAC_C1II	4.17	278.00	93.71
14	MAC_C2I	3.87	258.00	86.97
15	MAC_C2II	4.21	280.67	94.61
16	MAC_C2III	4.32	288.00	97.08
17	MAC_C3I	4.36	290.67	97.98
18	MAC_C3II	4.23	282.00	95.06
19	MAC_C3III	4.11	274.00	92.36
20	MAC_C4I	4.09	272.67	91.91
21	MAC_C4II	4.21	280.67	94.61
22	MAC_C4III	4.71	314.00	105.84
23	MAC_D1I	2.55	170.00	57.30
24	MAC_D1II	2.17	144.67	48.76
25	MAC_D3I	2.16	144.00	48.54
26	MAC_D3II	2.60	173.33	58.43
27	MAC_D4I	2.13	142.00	47.87
28	MAC_D4II	2.15	143.33	48.31
29	MAC_E1I	2.08	138.67	46.74
30	MAC_E1II	2.03	135.33	45.62
31	MAC_E3I	2.07	138.00	46.52
32	MAC_E3II	1.29	86.00	28.99
33	MAC_E4I	2.12	141.33	47.64
34	MAC E4II	2.38	158.67	53.48

Table 3	
Weight and percentage of MAC yield	l (%)

Notes: A = IFP (Iron Powder); B = I30B (γ -Fe₂O₃); C = α -Fe₂O₃; D = FeCl₂; E = FeSO₄; 1 = Control; 2 = D350; 3 = SDS; 4 = CA; I = Non-sonicated; II = Ultrasonic bath; III = Sonicated.

Methylene Blue Test

The behavior of methylene blue and iodine test were closely related to surface area and pores volume of the carbon sample. The parameter used in methylene blue test were the time taken by MAC to decolorize methylene blue solution (contact time) and the time taken by MAC fully attached to magnet (magnetic contact time). For methylene blue test, the selected MAC samples were from group A, B, D and E. Group C was not selected because group C did not contain any magnetism which was considered as lowest quality if compared with others. Table 4 shows the contact time and the magnetic contact time between the MAC sample and 2 ppm methylene blue. Additionally, the absorbance value had been recorded that measured at 668 nm by using UV-VIS. Besides, the test had been done triplicates and the average result was calculated. The results showed the contact time, magnetic contact time of the MAC form by using the iron powder (IFP) and treated with different chemical (control, D350, SDS and citric acid) and sonicated way (non-sonicated, ultrasonic bath and sonicated). The sample that achieved the shortest contact time was MAC A3I which recorded 5.38 s and on the other hand the longest contact time recorded was by MAC A2III which was 156.73 s. This explained that MAC A3I applied shorter time to decolorize the methylene blue whereas MAC A2III applied the longest time. For the magnetic contact time, the shortest time, 17.43 s recorded by MAC A2III and the longest time, 40.87 s recorded by MAC A2I. The result clarifies that some MAC have good methylene blue adsorption but weak magnetism. The magnetism of the MAC sample was affected by different types of iron powder used and the treated chemical.

Group B was the MAC prepared by using gamma phase of ferric oxide, γ -Fe₂O₃ (I30B) and treated by different chemical (control, D350, SDS and citric acid) and sonicated way (non-sonicated, ultrasonic bath and sonicated). Based on Table 4, MAC_B3I had recorded the shortest contact time which was 3.90 s whereas on the other hand MAC_B2I had recorded the longest contact time, 50.54 s. Furthermore, for the longest and shortest magnetic contact time, MAC_B2III and MAC_B1I recorded were 6.283 s and 71.87 s respectively.

The MAC from group D was carried out by corporate ferrous chloride (FeCl₂) with ferric chloride (FeCl₃). For group D, MAC_D3I and MAC_D1I had recorded the shortest and longest contact time when dissolved in methylene blue solution. Moreover, for the magnetic contact time, MAC_D3I displayed the best magnetism and conversely MAC_D1I displayed the worst magnetism among category D. The time recorded were 28.64 s and 35.91 s respectively. MAC_E which the group E was MAC that were manufactured by mixing the ferric chloride (FeCl₂) with ferrous sulphate (FeSO₄). Lastly for treatment Group E, the results showed that MAC_E3I and MAC_E4II had presented the shortest and longest contact time when they dissolved in methylene blue solution. The time recorded were 3.73 s and 4.54 respectively which also explained that MAC_E3I was more effective

No Sample		Contact	ct Magnetic Contact Absorbance Value		Percentage of
		Time (s)	Time (s)(at wavelength of 668 nm)		dye removal (%)
1	MAC_A1I	6.857	38.537	0.064 ± 0.004	81.009
2	MAC_A1II	10.713	30.743	0.060 ± 0.006	82.196
3	MAC_A2I	15.717	40.870	0.010 ± 0	97.033
4	MAC_A2II	32.283	31.380	0.102 ± 0.002	69.733
5	MAC_A2III	156.733	17.430	$0.194{\pm}0.0005$	42.433
6	MAC_A3I	5.357	35.140	0.013 ± 0.0011	96.142
7	MAC_A3II	7.427	33.420	0.004 ± 0.0005	98.81
8	MAC_A3III	10.437	30.310	0.003 ± 0.0005	99.110
9	MAC_A4I	21.107	19.790	0.009 ± 0.0005	97.329
10	MAC_A4II	11.883	35.110	$0.015{\pm}\ 0.001$	95.549
11	MAC_A4III	19.643	19.530	$0.011{\pm}\ 0.001$	96.736
1	MAC_B1I	5.533	71.867	$0.017 {\pm} 0.0005$	94.955
2	MAC_B1II	4.620	55.490	0.014 ± 0.0005	95.846
3	MAC_B2I	50.540	10.723	0.080 ± 0.0005	76.261
4	MAC_B2II	37.327	13.197	0.036±0	89.318
5	MAC_B2III	44.497	6.283	$0.118 {\pm} 0.0015$	64.985
6	MAC_B3I	3.903	54.997	$0.005 {\pm} 0.001$	98.516
7	MAC_B3II	4.040	53.527	0.009 ± 0.0005	97.329
8	MAC_B3III	3.953	45.783	0.003 ± 0.0005	98.110
9	MAC_B4I	5.737	49.437	0.030 ± 0	91.098
10	MAC_B4II	3.953	42.993	0.019 ± 0.0005	94.362
11	MAC_B4III	5.870	50.910	0.007 ± 0	97.923
1	MAC_D1I	4.727	30.070	0.049 ± 0.0005	85.460
2	MAC_D1II	3.917	35.910	$0.082{\pm}0.001$	75.668
3	MAC_D3I	3.600	28.640	0.067 ± 0	80.119
4	MAC_D3II	4.423	34.120	0.098 ± 0.0005	70.920
5	MAC_D4I	3.803	35.270	0.055 ± 0.0005	83.680
6	MAC_D4II	4.183	34.510	0.059 ± 0.0005	82.493
1	MAC_E1I	4.000	48.640	0.089 ± 0.0017	73.591
2	MAC_E1II	4.533	46.680	$0.074 {\pm} 0.001$	78.042
3	MAC_E3I	3.733	47.450	0.060 ± 0.0005	82.196
4	MAC_E3II	4.343	41.170	0.085 ± 0.0005	74.777
5	MAC_E4I	4.183	54.290	0.103 ± 0.00153	69.436
6	MAC_E4II	4.540	52.220	0.055 ± 0.0005	83.680

Table 4	
Methylene blue (2 ppm) test using different treatments	

Where n=3 for triplicate test of methylene blue test. Notes: A = IFP (Iron Powder); B = I30B (γ -Fe₂O₃); $D = FeCl_2$; $E = FeSO_4$; 1 = Control; 2 = D350; 3 = SDS; 4 = CA; I = Non-sonicated; II = Ultrasonic bath; III = Sonicated

if compared with MAC_E4II. Additionally, MAC_E3II and MAC_E4I had recorded 41.17 s and 54.29 s as the shortest and longest magnetic contact time. Table 5 showed the contact and magnetic contact time of the CMAC and AC. The result showed that the CMAC recorded the contact time of the sample in methylene blue solution as 4.13 s and 50.81 s as the magnetic contact time. On the other hand, due to AC not containing magnetism, the AC only recorded 9.58 s for the contact time of sample in methylene blue solution.

Table 5Methylene blue (2 ppm) test

No	Sample	Contact Time (s)	Magnetic Contact Time (s)		Absorbance Value (668 nm)
		Average	Average	Average	Percentage of dye removal (%)
1	CMAC	4.127	50.810	0.098	70.920
2	AC	9.580	-	0.244	27.596

Iodine Test

Iodine test for selected MAC sample were calculated by using Equation 3 to 5. The iodine number of the selected sample is shown in Table 6. The sample was selected based on the highest and the lowest of the percentage of dye removal from methylene blue test.

 Table 6

 Iodine number for selected MAC sample

No	Sample	Iodine Value (mg/g)
1	45µm AC	925.840
2	63 µm AC	874.948
3	CMAC	657.301
4	MAC_A2III	821.627
5	MAC_A3II	652.606
6	MAC_B2III	368.710
7	MAC_B3I	275.069
8	MAC_D1I	719.862
9	MAC_D3II	737.419
10	MAC_E4I	795.945
11	MAC_E4II	782.240

Notes: A = IFP (Iron Powder); B = I30B (γ -Fe₂O₃); $D = FeCl_2$; $E = FeSO_4$; 1 = Control; 2 = D350; 3 = SDS; I = Non-sonicated; II = Ultrasonic bath; III = Sonicated

Iodine test was used to determine the surface area of the MAC and AC. Additionally, iodine test also act as a measure of micropore content of AC by observing the iodine adsorption from the solution. As shown in Tables 3 and 4, there are comparison between

AC and MAC. At first, the comparison between 45 µm AC and 63 µm AC was made. The result clearly showed that the iodine number of 45 µm AC was higher than 63 µm AC. 45 μm AC recorded 925.84 mg/g whereas 63 μm AC recorded 874.95 mg/g of iodine number. The higher iodine number indicates a higher degree of activation (Machrouhi et al., 2019). Besides, higher iodine number indicates that the 45 µm AC contain a larger porosity and higher number of porosity (Arslanoglu, 2019). The result showed that 45 µm AC had higher quality compared to 63 µm AC. The iodine test for MAC was also calculated for comparison. For group A, MAC A3II recorded a lower iodine adsorption number, 652.606 mg/g which indicated it was better compared to MAC A2III that recorded higher iodine adsorption number, 821.63 mg/g (Figure 2). Next, for the comparison of other group B, D and E, the more qualified MAC are MAC B3I, MAC D1I and MAC E4II. The iodine adsorption number recorded are 275.07 mg/g, 719.86 mg/g and 782.24 mg/g respectively. MAC B3I recorded the lowest iodine adsorption number which indicated that the surface of MAC B3I are mostly covered by iron ion (Deng et al., 2010). The iodine adsorption number of MAC are much lower compared to AC because of the surface of MAC were mostly covered by iron ion depending on the type of magnetic agent used. Besides, this further explained that MAC A3II having a lower iodine adsorption number had a smaller number of micropores and the surface of carbon were mostly covered by iron ion.



Figure 2. Process flow on the synthesis of magnetic activated carbon treated with sodium dodecyl sulphate

Characterization of Magnetic Activated Carbon

Scanning Electron microscope. Scanning electron microscopy analysis is an analytical technique to study the physical properties of the carbon. This analysis was mainly to illustrate the pore structure or the morphology of the carbon sample. Figure 3 depicts the



Figure 3. Micrograph of carbon material at magnification of 1000×: (a) AC; (b) CMAC; (c)MAC_A3II; and (d) MAC_B2III

micrograph and image of the chosen sample under the magnification of 1000x. The sample were chosen based on the best condition on iodine test and methylene blue test and samples chosen were the AC, CMAC, MAC A3II and MAC B2III. Figure 3 shows the micrograph related to AC. As seen on the micrograph, there were abundant pores developed on the AC (1000x). The pores were randomly scattered on the surface of the activated carbon with various size, diameter, and width of external pores. Besides, there was also some white particles spotted on the surface of the carbon. This might be due to the impurities that was not washed completely during the washing process (Anyika, et al., 2017). The micrograph of the AC showed many micropores and mesopores compared to other samples. This was certainly proof with the iodine number that are higher if compared to other samples. The micrographs of CMAC, MAC A3II and MAC B2III are depicted in Figure 3 b, c, and d, respectively. The micrograph of all MAC sample does not show obvious pores if compared to activated carbon. This is because the iron powder used have blocked the pores of the MAC (Suresh et al., 2017). Furthermore, the carbon surface of CMAC showed a homogeneous surface whereas micrograph of MAC A3II and MAC B2III showed that the particle size was not homogeneous.

The micrograph of MAC_A3II displayed bigger and uneven size of particle compared to CMAC. In addition, MAC_A3II also showed some needle like structure which represented the presence of ferrous oxide. Moreover, MAC_B2III contained less greyish layer compared to the others. The greyish layer might represent some of the chemical that added to strengthen the magnetism of the MAC. Next, there were some white spots scattered around the surface of each magnetized sample. The white spot was believed to act as the silica particles and MAC_B2III contained more white spots because of the addition of D350 chemical that were contained in silica particles (Wang, et al., 2017).

X-Ray Diffraction (XRD). X-ray diffraction is the analytical technique that mainly identify the crystalline phase of the sample tested. The selected samples were analyzed by using X-ray diffraction analysis as shown on Figure 4. Diffractogram of AC showed some intense

Synthesis of MAC Treated With SDS



Figure 4. XRD diffractogram of AC, CMAC, MAC_A3II and MAC_B2III (Note: A = IFP (iron powder); B = I30B (γ -Fe₂O₃); 2 = D350; 3 = SDS; II = ultrasonic bath; III = Sonicate)

peaks which assigned to the presence of carbon at $20\ 26.54^\circ$, 42.42° , 59.74° , and 87.55° at intensity of 596.95, 483.38, 181.95 and 181.85, respectively. Diffractogram of AC shows that AC contained 47.8% of graphite compound. Diffractogram of CMAC had shown the main composition was iron oxide at 90.3%. The most important structure in MAC was the iron compound that leads to the magnetism of the AC. The diffractogram had shown a few peaks which matched to ferric oxide at $20\ 30.54^\circ$, 35.71° and 62.80° at the intensity of 418.86, 1000.00, and 456.86, respectively. The presence of iron oxides indicates that the CMAC contains magnetism. Furthermore, diffractogram of MAC_A3II displayed the many peaks that related to ferrous oxide. There were some intense peaks that matched to ferric oxide at $20\ 30.26^\circ$ and 35.68° at the intensity of 394.73 and 853.49, respectively. Diffractogram of MAC_B2III shows some peaks at $20\ 35.55^\circ$ and 63.96° that matched to maghemite and carbon whereas $20\ 26.54^\circ$ that matched to carbon.

CONCLUSION

This study revealed that MAC_A3II had presented a better characteristic compared with other adsorbent samples, CMAC and the AC. MAC_A3II produced by impregnated the iron powder with SDS as the treated chemical and sonicated by using ultrasonic bath. Moreover, MAC_A3II performed better methylene blue adsorption by achieving 98.81% of the removal percentage on 2 ppm methylene blue solution. The iodine test of MAC_A3II, 626.61 mg/g was lower than the AC which was 925.84 mg/g. This showed that AC were highly porous and had larger surface area. On the other hand, MAC_A3II had lower iodine number because the iron had blocked the surface of the carbon. The SEM analysis clearly showed that AC contained many micropores and mesopores if compared with MAC. Besides, for SEM analysis of MAC, the micropores and mesopores were very hard to identified as the pores were blocked by the iron or silicone atom. From the analysis it can be concluded that MAC_A3II presented better qualities and characteristics on the methylene blue adsorption and magnetism if compared to AC and CMAC. MAC_A3II prepared by using the iron powder and treated by adding SDS, was proven as the best qualities of MAC. The presence of iron oxide had been verified by using XRD analysis.

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